



人粒细胞—巨噬细胞集落刺激因子定量分析酶联免疫检测试剂盒

本试剂盒仅供科研使用。用于体外定量检测人血清、血浆或细胞培养上清液中的 GM-CSF 浓度。**使用前请仔细阅读说明书并检查试剂盒分是否完整。**如有产品包装破损或质量投诉，请在收到货一个月之内联系我们。

GM-CSF 简介:

粒细胞—巨噬细胞集落刺激因子 (GM-CSF) 最早作为粒细胞—巨噬细胞前体细胞体外增殖的刺激因子来命名的。它可由包括 T 细胞、B 细胞、巨噬细胞、肥大细胞、内皮细胞和成纤维细胞等细胞对炎症刺激而产生。人的 GM-CSF 是由包括 17 个信号肽的前体裂解为包括 124 和 127 氨基酸的成熟蛋白。

GM-CSF 是红细胞系、巨核细胞和嗜酸粒细胞等的祖细胞的生长因子。对成熟的造血系细胞来说，GM-CSF 是一种生长支持因子，是粒细胞、单核/巨噬细胞、嗜酸性细胞的功能活化因子。GM-CSF 也被发现对非造血系细胞具有重要的作用，如它能介导人的内皮细胞的的迁移和增生等。

在人的前列腺癌、严重性创伤、甲状腺癌、严重的粘膜炎、真菌感染、AIDS、急性淋巴性白血病、恶性血液病、感染、肺癌等患者的血液中都检测出有相关的 GM-CSF 的波动。

检测原理:

本试剂盒采用双抗体夹心ELISA法检测样本中GM-CSF的浓度。GM-CSF捕获抗体已预包被于酶标板上，当加入标本或参考品时，其中的GM-CSF会与捕获抗体结合，其它游离的成分通过洗涤的过程被除去。当加入生物素化的抗人GM-CSF抗体后，抗人GM-CSF抗体与GM-CSF接合，形成夹心的免疫复合物，其它游离的成分通过洗涤的过程被除去。随后加入辣根过氧化物酶标记的亲合素。生物素与亲合素特异性结合，亲合素连接的酶就会与夹心的免疫复合物连接起来；其它游离的成分通过洗涤的过程被除去。最后加入显色剂，若样本中存在GM-CSF将会形成免疫复合物，辣根过氧化物酶会催化无色的显色剂氧化成蓝色物质，在加入终止液后呈黄色。通过酶标仪检测，读其450nm处的OD值，GM-CSF浓度与OD₄₅₀值之间呈正比，通过参考品绘制标准曲线，对照未知样本中OD值，即可算出标本中GM-CSF浓度。

人GM-CSF定量分析酶联免疫检测试剂盒组成:

组分	规格 (96T/48T)
人GM-CSF预包被板	12条/6条
样本分析缓冲液	5ml/3ml
标准品稀释液	10ml/5ml
人GM-CSF标准品	2支/1支(冻干)*
人GM-CSF生物素化抗体	10ml/5ml
亲和素连接的HRP酶	10ml/5ml
浓缩洗涤液 20×	30ml/15ml
TMB底物	10ml/5ml
中止液	5ml/3ml
封板胶纸	3/2张
说明书	1份

标本收集:

- 标本的收集请按下列流程进行操作：
 - 细胞上清标本离心去除悬浮物后即可；
 - 血清标本应是自然凝固后，取上清，避免在冰箱中凝固血液；
 - 血浆标本，推荐用EDTA的方法收集若待测样本不能及时检测，
 - 标本收集后请分装，冻存于-20℃，避免反复冻融。
- 血清标本不应添加任何防腐剂或抗凝剂；
- 标本应清澈透明，检测前样本中如有悬浮物应通过离心去除。



4. 请勿使用溶血，高血脂或污染的标本检测，否则结果将不准确。

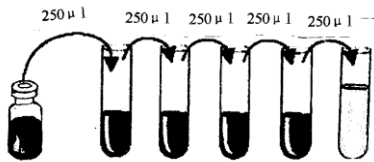
注：人血清或血浆样本请用样本分析缓冲液做倍比稀释后再检测。

注意事项：

1. 试剂盒请保存在2~8℃。
2. 浓缩洗涤液因在低温下可能有结晶，请水浴加热使结晶完全溶解后再配制工作液。
3. 标准品复溶加样后，剩余部份请丢弃。
4. 底物请勿接触氧化剂和金属。
5. 加样时，请及时更换枪头，避免交叉污染。
6. 严禁混用不同批号的试剂盒组份。
7. 充分混匀对保证反应结果的准确性很重要，在加液后请轻轻叩击边缘以保证混匀。
8. 室温反应，请严格控制在25~28℃。
9. 洗涤过程是至关重要的，洗涤不充分会使精确度下降并导致结果误差较大。
10. 试验中标准品和样本检测时建议作双复孔。
11. 加样过程中避免气泡的产生。
12. 血清和血浆标本的检测时，检测抗体的孵育时间应适当延长。

检测前准备工作：

1. 试剂盒自冰箱中取出后应置室温（25~28℃）平衡20分钟；每次检测后剩余试剂请及时于2~8℃保存。
2. 将浓缩洗涤液用双蒸水或去离子水稀释（1份加19份水）。
3. 如有5X标准品稀释液按所需用双蒸水或去离子水稀释（1份加4份水）。
4. 标准品：按标签复溶体积加入准品稀释液复溶使GM-CSF终浓度达到1000pg/ml，室温反应，请严格控制在25~28℃，静置10~15分钟后轻轻混悬（建议抽吸几次）待彻底溶解，用标准品稀释液倍比梯度稀释后依次加入检测孔中。（标准曲线取七个点，最高浓度为1000 pg/ml，标准品稀释液直接加入作为0浓度。）



洗涤方法：

自动洗板机或人工洗板：每孔洗涤液为300ul，注入与吸出间隔15-30秒。洗板5次。最后一次洗板完成后将板倒扣着在厚吸水纸上用力拍干。

实验过程需自备的材料：

1. 不同规格的加样枪及相应的枪头；
2. 酶标仪；
3. 自动洗板机；
4. 去离子水或双蒸水；

操作步骤：

1. 通过计算并确定一次性实验所需的板条数，取出所需板条放置在框架内，暂时用不到板条请放回铝箔袋密封，保存于4℃。
2. 建议设置本底校正孔，即空白孔，设置方法为该孔只加TMB显色液和终止液。每次实验均需做标准品对照并画出标准曲线。
3. 分别将标本或不同浓度标准品（100ul/孔）加入相应孔中，用封板胶纸封住反应孔，室温（25~28℃）孵育120分钟。对于血清或血浆标本，请加150 ul样本分析缓冲液后加50 ul标本，如稀释量大，请将样本与样本分析缓冲液等量加入，不足部分用标准品稀释液补充至100ul。
4. 洗板5次，且最后一次置厚吸水纸上拍干。
5. 加入生物素化抗体工作液（100ul/孔）。用封板胶纸封住反应孔，室温（25~28℃）孵育60分钟。
6. 洗板5次，且最后一次置厚吸水纸上拍干。
7. 加入亲和素连接的HRP酶工作液（100ul/孔）。用封板胶纸封住反应孔，避光室温（25~28℃）孵育20分钟。



8. 洗板5次，且最后一次置厚吸水纸上拍干。
9. 加入显色剂TMB100 μ l/孔，避光室温（25~28 $^{\circ}$ C）孵育20分钟。
10. 加入终止液50 μ l/孔，混匀后即刻测量OD450值。

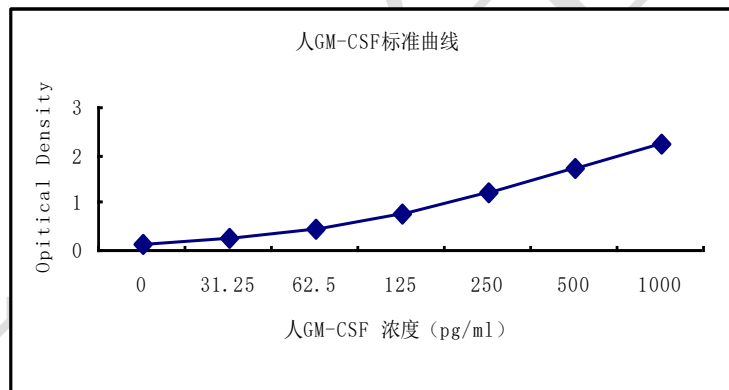
结果判断：

1. 复孔的值在20%的差异范围内结果才有效，复孔的值平均后可作为测量值。
2. 每个标准品或标本的OD值应减去本底校正孔的OD值。
3. 手工绘制标准曲线。以标准品浓度作横坐标，OD值作纵坐标，以平滑线连接各标准品的坐标点。通过标本的OD值可在标准曲线上查出其浓度。
4. 若标本 OD 值高于标准曲线上限，应适当稀释后重测，计算浓度时应乘以稀释倍数。

典型数值和参考曲线

浓度pg/ml	典型OD值1	典型OD值2	OD平均值
0	0.1258	0.0894	0.1076
31.25	0.3589	0.1949	0.2769
62.5	0.1598	0.7944	0.4771
125	0.8954	0.6958	0.7956
250	1.0247	1.3673	1.196
500	1.6853	1.7229	1.7041
1000	2.2471	2.2267	2.2369

人GM-CSF参考标准曲线



注意：本图仅供参考，应以同次试验标准品所绘标准曲线计算标本含量。

灵敏度，特异性和重复性：

1. 灵敏度：多次重复结果表明，最小检出量为12.5pg/ml。
2. 特异性：与人EGF, Epo, FGF acidic, FGF basic, G-CSF, HGF, IFN- γ IGF-I, IGF-II, IL-1, IL-2, IL-4, IL-6, IL-8, IL-10, IL-12, IL-13, KGF, LIF, M-CSF, MCP-1, SCF, TGF- α , TGF- β 1, VEGF等没有交叉反应。
3. 重复性：板内，板间变异系数均<10%。

参考文献：

1. Gasson, J. (1991) Blood 77:1131.
2. Costello, R.T. (1993) Acta Oncol. 32:403.
3. Crosier, K.E. et al. (1991) Proc. Natl. Acad. Sci. USA 88:7744.
4. Lopez, A.F. et al. (1990) J. Cell Physiol. 145:69.
5. Omori, F. et al. (1992) Biotherapy 4:147.
6. Nakamura, Y. et al. (1993) Am. Rev. Respir. Dis. 147:87.



ELISA Kit for the Quantitative Analysis of Human GM-CSF

The human GM-CSF ELISA (enzyme-linked immunosorbent assay) kit is used for detection of human GM-CSF in cell culture supernatants, human serum and plasma. **THE ELISA KIT IS FOR RESEARCH USE ONLY.** Please read this instruction manual carefully and check out the material provided before use, and you can contact with our company if any questions. You can enter our website or call us for other aim.

Introduction

Granulocyte-Macrophage Colony Stimulating Factor (GM-CSF) was initially characterized as a growth factor that can support the in vitro colony formation of granulocyte-macrophage progenitors. It is produced by a variety of cell types including T cells, B cells, macrophages, mast cells, endothelial cells and fibroblasts in response to cytokine or immune and inflammatory stimuli. human GM-CSF encode precursor proteins with a 17 amino acid signal peptide that is cleaved to form mature proteins with 124 and 127 amino acids.

GM-CSF is a growth factor for erythroid, megakaryocyte and eosinophil progenitors. On mature hematopoietic cells, GM-CSF is a survival factor for and activates the effector functions of granulocytes, monocytes/macrophages and eosinophils. GM-CSF has also been reported to have a functional role on non-hematopoietic cells. It can induce human endothelial cells to migrate and proliferate. Monitoring of GM-CSF has been found relevant in human prostate cancer, poorly healing wounds, thyroid carcinoma, severe mucositis, fungal infections, AIDS, bone marrow transplantation, renal cell carcinoma and acute lymphoblastic leukemia pulmonary inflammation, hematological malignancies, infection, lung cancer.

Principles of the Test

The kits is a solid sandwich enzyme-linked immunosorbent assay for detection of human GM-CSF. An anti-human GM-CSF monoclonal antibody has been absorbed onto the wells of the microtiter strips provided. Samples including specimens or standards were pipetted into wells. The human GM-CSF in specimens or standards would be captured by the coated antibody and the free others were removed by washing. The human GM-CSF biotin-conjugated antibody were added and binds to human GM-CSF captured by the first antibody, which formed a sandwich. Streptavidin-HRP would be added and binds to the biotin conjugated antibody, then free Streptavidin-HRP would be removed during a wash step. After this, substrate solution would be added and catalyzed by the HRP, and a coloured product is formed. The intensity of the colored product is used to calculate in proportion to the amount of human GM-CSF in the original specimen.

Materials provided with the kits:

reagent	96/48 Test Kit
Assay Buffer	5ml/3ml
Human GM-CSF Antibody-Coated Wells	12 strips/6 strips
Standard Diluent	10 ml/5ml
Human GM-CSF Standard	2/1 vial(s)
Human GM-CSF Detection Antibody	10ml/5ml
Streptavidin-HRP	10ml/5ml
Wash Buffer Concentrate 20x	30ml/15ml
TMB	10ml/5ml
Stop Solution	5ml/3ml
Plate Covers	3/2
Complete Instruction Manual	1

Specimen Collection

- Collecting specimen as following:
 - The particulate of the cell culture supernatants should be removed before use.
 - Serum was obtained from clot at room temperature.
 - Please collect plasma with EDTA.
 - Assay immediately or store samples at -20°C . Avoid free-thaw cycles.
- Antiseptic and anticoagulant should not appear in Serum samples.



3. Any particulate should be removed from samples before use.

4. Do not use grossly hemolyzed or lipemic samples.

Note: Strongly recommend that the serum and plasma samples should be diluted as doubling dilution before use.

Precautions for use:

1. Please store the Kit at 2~8°C.
2. Washing buffer concentrate may have crystal in low temperature, and you can melt it in water-bath before use.
3. Please discard the remains after use of the dissolved standard.
4. Avoid contact of substrate solution with oxidizing agents and metal.
5. Usage of disposable pipette tips avoid microbial contamination or cross-contamination of reagents or specimens.
6. Do not mix or substitute reagents with those from other lots or other sources.
7. To ensure the adequate mixture of added reagents, please tap gently the plate after the wells were filled with liquid.
8. Incubation temperature should be 25~28°C.
9. Wash step was crucial for whole assay process.
10. Duplicate wells of the same sample were recommended in assay process.
11. Avoid the foam while pour the liquid into wells.
12. For serum or plasma samples, the biotin-conjugated antibody should be incubated for at least 90 minutes.

Reagent Preparation

1. The reagents should be warmed up to room temperature before use. The remaining reagents must be resealed and put into refrigeration again as soon as possible.
2. Dilute 1ml of wash buffer concentrate into 19ml deionized or distilled water to work.
3. If you have a 5x standard diluent, please dilute it with double distilled water or deionized water.
4. Add standard diluent to the bottle according to the volume of the label and wait 15 minutes for complete dissolution. Incubation temperature should be 25~28°C. And in turn add the half concentration diluent by standard diluent.

Wash step:

Automated microplate washer or operating by pipette: Each well should be poured into 300ul wash buffer and soaked for 15 or 30 seconds, then aspirated, five times process were repeated. After the last wash, remove remaining wash buffer by aspirating. Invert the plate and blot it against clean paper towels.

Materials Required But Not Provided

1. pipettes and pipette tips
2. Microwell strip reader capable of reading at 450 nm (540 nm as optional reference wavelength)
3. automated microplate washer
4. Glass-distilled or deionized water

Assay procedure

1. The needed strips were put into the frame, the remains were returned into foil pouch and resealed.
2. Blank well were recommended, which only color reagent and stop solution be added. It is suggested that each testing with gradient density of standard for standard curve.
3. Add 100ul of standard or sample. Cover with the Plate Covers provided. Incubate for 2 hours at room temperature. If assay the serum sample, you should add 50μl assay buffer with 50μl sample into the wells, if the protein concentration is higher than the range of the Kit, add the same quantity assay buffer with the sample, the deficiency should be complemented with sample diluent to 100μl per well.
4. Five times wash process were repeated.
5. Add 100ul of detection antibody. Cover with the Plate Covers provided. Incubate for 1 hour at room temperature.
6. Five times wash process were repeated.
7. Add 100ul of Streptavidin-HRP. Cover with the Plate Covers provided. Luciferal incubation for 20 minutes at room temperature.
8. Five times wash process were repeated.
9. Add 100ul of TMB, Luciferal incubation for 20 minutes at room temperature.
10. Add 50ul of stop solution to each well, determine the optical density of each well within 10 minutes.

Calculation of Results

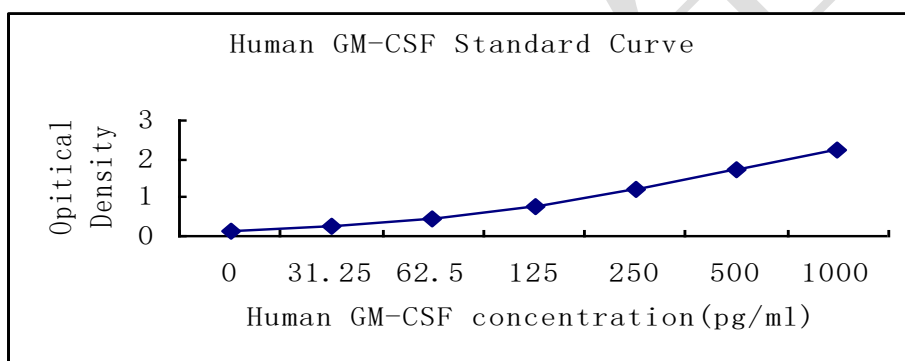


1. Duplicates should be within 20 per cent of the mean. Average absorbance values for each set of duplicate samples were used as detection results.
2. The blank absorbance values of subtract should be deducted.
3. Drawing a best fit curve through the points of graph. Draw the standard curve by plotting assayed OD value (on the Y axis) vs. concentration (on the X axis). The sample concentration was obtained based on its OD value founding in the standard concentration curve.
4. If the values obtained are not within the expected range of the standard, Samples should be dilute and assay again.

Typical Data and Standard Curve

concentration (pg/ml)	Typical data 1	Typical data 2	Average
0	0.1258	0.0894	0.1076
31.25	0.3589	0.1949	0.2769
62.5	0.1598	0.7944	0.4771
125	0.8954	0.6958	0.7956
250	1.0247	1.3673	1.196
500	1.6853	1.7229	1.7041
1000	2.2471	2.2267	2.2369

Human GM-CSF standard curve



Sensitivity, Specificity, Repeatability

Sensitivity: repeated assays were evaluated and the minimum detectable dose was 12.5pg/ml.

Specificity : No significant cross-reactivity or interference with human EGF,Epo,FGF acidic,FGF basic, G-CSF, HGF, IFN- γ , IGF-I ,IGF-II ,IL-1,IL-2,IL-4,IL-6,IL-8,IL-10,IL-12,IL-13,KGF,LIF,M-CSF,MCP-1,SCF,TGF- α ,TGF- β 1,VEGF

Repeatability: The coefficient of variation between wells or plates is less than 10 percent.

REFERENCES:

1. Gasson, J. (1991) Blood 77:1131.
2. Costello, R.T. (1993) Acta Oncol. 32:403.
3. Crosier, K.E. et al. (1991) Proc. Natl. Acad. Sci. USA 88:7744.
4. Lopez, A.F. et al. (1990) J. Cell Physiol. 145:69.
5. Omori, F. et al. (1992) Biotherapy 4:147.
6. Nakamura, Y. et al. (1993) Am. Rev. Respir. Dis. 147:87.